

## Maldistribution of Neurofilaments, Disease Pathogenesis, and Amyotrophic Lateral Sclerosis

Rodolphe Perrot, PhD<sup>1</sup> and Jean-Pierre Julien, PhD<sup>2</sup>

1. Post-doctoral Fellow; 2. Professor and Canada Research Chair, Department of Anatomy and Physiology, Research Center, Laval University Hospital Center

### Abstract

Accumulations of neuronal intermediate filaments (IFs) are a characteristic of many human neurodegenerative disorders, including amyotrophic lateral sclerosis (ALS). However, IF formation and the contribution of IF aggregates to the pathogenesis of ALS remain unclear. Here, we review the possible mechanisms underlying the formation of IF accumulations and the mouse models that have been used to investigate the role of IF proteins in ALS pathogenesis.

### Keywords

Neurofilaments, axonal transport, protein aggregates, amyotrophic lateral sclerosis, transgenic mice, superoxide dismutase, motor neurons

**Disclosure:** Rodolphe Perrot, PhD, is the recipient of a post-doctoral fellowship from the ALS Society of Canada. Jean-Pierre Julien, PhD, holds a Canada Research Chair in neurodegeneration. The authors have no conflicts of interest to declare.

**Received:** February 9, 2009 **Accepted:** September 4, 2009 **DOI:** 10.17925/USN.2010.05.02.30

**Correspondence:** Jean-Pierre Julien, PhD, Centre de Recherche du Centre Hospitalier Universitaire de Québec, Pavillon CHUL, 2705 Boulevard Laurier, Québec, G1V 4G2, Canada. E: jean-pierre.julien@crchul.ulaval.ca

Amyotrophic lateral sclerosis (ALS), also called Lou Gehrig's disease, is a late-onset progressive motor neuron disease first identified in 1869 by Jean-Martin Charcot. Its incidence is approximately two cases per 100,000, with a slightly higher prevalence in men. The progressive degeneration of motor neurons in ALS leads to neuron death and loss of muscle function. Patients become partially or totally paralyzed, but their cognitive functions usually remain unaffected. There is no cure for ALS and the disease is usually fatal within three to five years of the onset of symptoms. About 90% of ALS cases are sporadic, while approximately 10% are inherited in a dominant manner. Mutations in the gene coding for superoxide dismutase 1 (SOD1) account for 20% of all familial cases. To date, more than 140 mutations have been found in the *SOD1* gene, and transgenic mice overexpressing various *SOD1* mutants develop an ALS-like phenotype through a gain of unknown toxic properties.

### Intermediate Filament Alterations in Amyotrophic Lateral Sclerosis

The neuronal cytoskeleton is composed of three interconnected structures: actin microfilaments, microtubules, and intermediate filaments (IFs). Neurofilaments (NFs) are the major IFs present in adult neurons and their expression is restricted to neuronal cell types. Neurons express differentially several IF proteins depending on their stage of development or their localization in the nervous system: nestin (200kDa), NF triplet proteins (NFL [light, 68kDa], NFM [medium, 160kDa], and NFH [heavy, 205kDa]),  $\alpha$ -internexin (66kDa), peripherin (57kDa), and synemin (41kDa).<sup>1</sup>

One of the pathological features of both sporadic and familial ALS is the presence in motor neurons of axonal spheroids and perikaryal accumulations comprising NFs and/or peripherin.<sup>2</sup> NFs in perikaryal aggregates are extensively phosphorylated, while this occurs normally only within the axon.<sup>3</sup> A 70% decrease in levels of NFL messenger RNA (mRNA) in degenerating neurons of ALS was reported concomitantly with a decrease of  $\alpha$ -internexin or peripherin mRNA levels.<sup>4</sup> It is also interesting to note the presence of high NFL levels and auto-antibodies against NFL in the cerebrospinal fluid of ALS patients.<sup>5,6</sup>

Codon deletions or insertions in the lysine-serine-proline (KSP) repeat motifs of NFH have been identified in a small number of patients with sporadic ALS,<sup>7-9</sup> and are being viewed as a risk factor for the disease. However, two other studies failed to identify variants in the NF genes linked to sporadic and familial ALS,<sup>10,11</sup> suggesting that mutations in the NF genes are not a systematic common cause of ALS. By contrast, several mutations in the *NEFL* gene have been linked to Charcot-Marie-Tooth disease.<sup>12-15</sup> A frameshift deletion in the peripherin gene has also been discovered in a sporadic ALS case.<sup>16</sup>

Neuronal IF abnormalities in ALS may also occur as a result of post-translational protein modifications. Indeed, advanced glycation end-products were detected in NF aggregates of motor neurons in familial and sporadic ALS.<sup>17</sup> Moreover, Crow et al.<sup>18</sup> showed that SOD1 can catalyze nitration of tyrosines by peroxynitrite in the rod and head domains of NFL. However, no significant changes were detected in the nitration of NFL isolated from cervical spinal cord tissue of sporadic ALS cases.<sup>19</sup>

**Table 1: Toxic Agents that Cause the Accumulation of Neurofilaments**

Toxic Agents	Effects	References
2,5-hexanedione (HD)	<ul style="list-style-type: none"> <li>• Causes direct <math>\gamma</math>-diketone modification of NFs involving pyrrolation of <math>\epsilon</math>-amino groups on NF lysyl residues and possibly secondary autoxidation of the pyrrole rings with creation of covalent NF–NF cross-links</li> <li>• Impairs the axonal transport of NFs</li> <li>• The aggregation of NFs occurs in the distal region</li> </ul>	55
3,4-dimethyl-2,5-hexanedione (DMHD)	<ul style="list-style-type: none"> <li>• Impairs the axonal transport of NFs</li> <li>• The aggregation of NFs occurs in the proximal region</li> <li>• 20–30 times more potent as a neurotoxicant than 2,5-hexanedione</li> </ul>	55
$\beta$ - $\beta'$ -iminodipropionitrile (IDPN)	<ul style="list-style-type: none"> <li>• Segregates MTs from NFs</li> <li>• The aggregation of NFs occurs in the proximal region of axons through abnormal cross-linking of hyperphosphorylated NFs</li> <li>• NF-rich large-caliber axons are the most affected</li> <li>• NFH protein is a key mediator of IDPN-induced axonopathy</li> </ul>	56–60
Acrylamide	<ul style="list-style-type: none"> <li>• Inhibits the axonal transport of NFs</li> <li>• Alters NF protein gene expression</li> <li>• Increases <math>\text{Ca}^{2+}</math>/CaM-dependent phosphorylation of NF triplet proteins</li> <li>• Decreases NF degradation</li> <li>• NFs accumulate in proximal axons</li> </ul>	61–64
Aluminium	<ul style="list-style-type: none"> <li>• Inhibits NF degradation and dephosphorylation</li> <li>• Reduces the assembly of newly synthesized NF subunits into NF</li> <li>• Causes the formation of NF tangles in neuronal perikarya and proximal parts of dendrites</li> </ul>	65–68
Arsenic	<ul style="list-style-type: none"> <li>• Decreased levels of NFs are observed in sciatic nerve following arsenic exposure</li> <li>• Decreases NF transport and induces the perikaryal accumulation of phosphorylated NFs</li> </ul>	69, 70
Carbon disulfide	<ul style="list-style-type: none"> <li>• Causes intramolecular and intermolecular crosslinking of NF proteins</li> <li>• Disrupts the axonal transport of NFs</li> <li>• Leads to the gradual accumulation of NFs proximal to the nodes of Ranvier</li> </ul>	71, 72
Lead	<ul style="list-style-type: none"> <li>• Decreases amount and phosphorylation of NFM within the axons connecting auditory nuclei in the avian brainstem</li> <li>• Increases NF phosphorylation and neuritic beading within the murine auditory brainstem</li> </ul>	73, 74
Tri- <i>o</i> -cresyl phosphate	<ul style="list-style-type: none"> <li>• Decreases expression of NF triplet proteins</li> <li>• Increases phosphorylation of NF triplet proteins</li> </ul>	75, 76

MT = microtubule; NF = neurofilament; NFH = heavy NF.

### Possible Mechanisms Leading to the Formation of Intermediate Filament Aggregates in Amyotrophic Lateral Sclerosis

Several factors can potentially induce the accumulation of NFs in neurodegenerative diseases, including dysregulation of NF gene expression, NF mutations, defective axonal transport, abnormal post-translational modifications, and proteolysis. Toxic agents are also responsible for NF aggregations (see *Table 1*).

In the case of ALS, the mechanisms governing the formation of IF aggregates are still not clearly established. There is evidence that IF accumulations could result from defects of axonal transport or from abnormal stoichiometry of IF proteins. Perturbations of NF axonal transport is one of the earliest pathological changes seen in several transgenic mouse models of ALS.<sup>20–22</sup>

Phosphorylation changes might be involved. Abnormal phosphorylation of NFs can affect their axonal transport and lead to their accumulation in cell bodies and in proximal axons. Many studies support the view that the rate of NF transport is inversely correlated to their phosphorylation state (for a review see reference 1). The premature phosphorylation of NF tail domains in motor neuron cell bodies could directly mediate their accumulation in this region. Glutamate excitotoxicity, another pathogenic process in ALS, may

induce abnormal phosphorylation of NFs. Treatment of primary neurons with glutamate activates members of the mitogen-activated protein kinase (MAPK) family that phosphorylate NFs with ensuing slowing of their axonal transport.<sup>23</sup> In addition, glutamate leads to caspase cleavage and activation of protein kinase N1 (PKN1), an NF head-rod domain kinase.<sup>24</sup> This cleaved form of PKN1 disrupts NF organization and axonal transport.

Finally, excitotoxicity mediated by non-N-methyl-D-aspartic acid (NMDA) receptors is also associated with the aberrant co-localization of phosphorylated and dephosphorylated NF proteins.<sup>25</sup> Inhibition of Pin1 was suggested as a possible therapeutic target to reduce pathological accumulation of phosphorylated NFs.<sup>26</sup> Pin1 is a prolyl isomerase that selectively binds to phosphorylated proline-directed serine/threonine residues in target proteins and isomerizes *cis* isomers to more stable *trans* configurations. Pin1 associates with phosphorylated NFH in neurons and is co-localized in aggregates found in the spinal cord of patients with ALS.<sup>26</sup> The inhibition of Pin1 reduces glutamate-induced perikaryal accumulation of phosphorylated NFH.

Alterations of the anterograde or retrograde molecular motors may also be responsible for the aggregation of IFs. Mutation of dynein or p150<sup>glued</sup>,<sup>27</sup> overexpression of dynamitin (the sub-unit of the dynein–

**Table 2: Mouse Models of Amyotrophic Lateral Sclerosis with Alterations of the Intermediate Filament Network**

Mice	Modifications of Neuronal Cytoskeleton	Modifications of Axonal Transport	Other Pathological Changes
<b>Cytoskeletal Components</b>			
hNFH <sup>21,34,35</sup>	<ul style="list-style-type: none"> <li>Abnormal accumulation of NFs in the perikaryon and proximal axons of spinal motor neurons</li> <li>Reduced number of normal NF structure in motor axons</li> </ul>	Slowing of NF, actin, and tubulin transport in sciatic nerve at 2 and 3 months of age	<ul style="list-style-type: none"> <li>Progressive motor dysfunction and weakness</li> <li>Reduction of bodyweight with aging</li> <li>Loss of motor axons at 2 years of age</li> </ul>
hNFM (high level) <sup>77</sup>	<ul style="list-style-type: none"> <li>Neurofilamentous accumulations in ventral horn motor neurons</li> <li>Increased NF density in motor axons</li> </ul>	Not determined	<ul style="list-style-type: none"> <li>Hind limb paralysis</li> <li>Axonal loss in (77) ventral motor roots</li> </ul>
NFL(Pro) mutant mice <sup>42</sup>	<ul style="list-style-type: none"> <li>Perikaryal and proximal axonal NF accumulations in motor and sensory neurons</li> <li>25–50% decrease of NF content in sciatic nerve</li> </ul>	Not determined	<ul style="list-style-type: none"> <li>Massive degeneration of spinal motor neurons but no degeneration of sensory neurons</li> <li>Severe neurogenic atrophy of skeletal muscle resulting in severe fore- and hind-limb weakness</li> <li>Early death</li> </ul>
NFL <sup>-/-78-81</sup>	<ul style="list-style-type: none"> <li>Perikaryal accumulation of NFM and NFH</li> <li>Scarcity of IFs and increased MT density in axons</li> </ul>	Normal rate of axonal transport of NFM in optic nerve	<ul style="list-style-type: none"> <li>20% loss of motor axons</li> <li>Delayed axonal regeneration</li> <li>Mild sensory motor dysfunction</li> </ul>
Mouse NFL overexpressor <sup>32,82</sup>	<ul style="list-style-type: none"> <li>Massive accumulation of NFs in perikarya, axons and dendrites from motor neurons and in some dorsal root ganglion neurons</li> <li>Twofold increased NF density in large myelinated axons in ventral root</li> </ul>	Not determined	<ul style="list-style-type: none"> <li>Loss of motor axons</li> <li>Severe skeletal muscle atrophy</li> <li>Death before 3 weeks of age</li> </ul>
Peripherin overexpressor <sup>36-38,83</sup>	<ul style="list-style-type: none"> <li>Age-dependant IF aggregates in perikarya and axons</li> </ul>	Slowing of NF transport in sciatic nerve at 6 months of age	<ul style="list-style-type: none"> <li>35% loss of spinal motor neurons at 28 months</li> <li>The onset of peripherin-mediated disease is precipitated by a deficiency of NFL</li> </ul>
Short tau overexpressor <sup>84</sup>	<ul style="list-style-type: none"> <li>NF inclusions in spinal cord neurons and proximal axons</li> <li>Reduced MT density at 12 months in ventral roots but unchanged NF density</li> </ul>	Reduced anterograde fast axonal transport in the ventral root axons	<ul style="list-style-type: none"> <li>20% loss of motor axons at 12 months</li> <li>Progressive motor weakness</li> </ul>
<b>MT-based Transport Components</b>			
Dynamitin overexpressor <sup>28</sup>	<ul style="list-style-type: none"> <li>Axonal IF swellings</li> </ul>	Delay in retrograde transport	<ul style="list-style-type: none"> <li>Decreased strength and endurance</li> <li>Loss of motor neurons and denervation of muscle</li> </ul>
KIF5A <sup>-/-29</sup>	<ul style="list-style-type: none"> <li>NF and peripherin accumulations in DRG sensory neuron cell bodies</li> </ul>	NF transport defects	<ul style="list-style-type: none"> <li>Sensory neuron degeneration but no degeneration of motor neurons</li> </ul>
BICD2-N overexpressor <sup>31</sup>	<ul style="list-style-type: none"> <li>Giant proximal NF swellings in motor axons</li> </ul>	Impaired retrograde trafficking	<ul style="list-style-type: none"> <li>Golgi fragmentation in motor neurons</li> <li>No motor neuron degeneration and motor abnormalities</li> </ul>
Dynein mutations <sup>27,85</sup>	<ul style="list-style-type: none"> <li>Perinuclear NF inclusions</li> </ul>	Impaired retrograde trafficking	<ul style="list-style-type: none"> <li>Progressive motor degeneration</li> </ul>
Mutant human p150 <sup>glued86</sup>	<ul style="list-style-type: none"> <li>Massive accumulation of NFs in proximal motor axons</li> </ul>	Defects in vesicular transport in cell bodies of motor neurons	<ul style="list-style-type: none"> <li>Tremors, weakness accompanied by muscle wasting in hind limbs</li> <li>Loss of large motor neurons</li> </ul>
<b>ALS-linked SOD1 Mutations</b>			
SOD1 mutant overexpressor <sup>20,22,44,45</sup>	<ul style="list-style-type: none"> <li>NF accumulations in cell bodies and axonal compartment of motor neurons</li> </ul>	Perturbations of slow and fast axonal transport	<ul style="list-style-type: none"> <li>70% loss of spinal motor neurons</li> </ul>

DRG = dorsal root ganglia; IF = intermediate filament; MT = microtubule; NF = neurofilament; NFH = heavy NF; NFL = light NF; NFM = medium NF.

dynactin complex that dissociates cargo from dynein),<sup>28</sup> and absence of kinesin heavy chain isoform 5A (KIF5A)<sup>29</sup> induce NF accumulation in mice. Recent studies suggest that inhibition of retrograde transport is more prone to causing accumulation of NFs than is inhibition of anterograde transport. The inhibition of dynein by increasing the level of dynamitin induces aberrant focal accumulation of NFs within axonal neurites, whereas inhibition of kinesin inhibits anterograde transport but does not induce similar focal aggregations.<sup>30</sup> Similarly,

the neuron-specific expression of bicaudal D2 N-terminus (BICD2-N), a motor-adaptor protein, impairs dynein–dynactin function, causing the appearance of giant NF swellings in the proximal axon.<sup>31</sup>

Modification in NF stoichiometry was also proposed to induce accumulation of NFs. The overexpression of different NF sub-units in mice provokes the formation of NF aggregates.<sup>32–34</sup> Remarkably, the motor neuron disease caused by excess human NFH (hNFH) can be

rescued by overexpression of hNFL in a dosage-dependent fashion.<sup>35</sup> Overexpression of peripherin provokes late-onset motor neuron degeneration in transgenic mice.<sup>36,37</sup> The motor neuron loss is preceded by axonal transport defects and formation of axonal spheroids.<sup>38</sup> Interestingly, NFH overexpression abolished the formation of axonal spheroids that might block transport. These findings illustrate the importance of IF protein stoichiometry in the formation, distribution, and toxicity of neuronal IF inclusions.

In ALS, there is a 70% decrease in levels of NFL mRNA observed in degenerating motor neurons.<sup>4</sup> This could be due in part to modification in the stability of their mRNA. Ge et al.<sup>39</sup> have shown that ALS-linked SOD1 mutant proteins bind to and destabilize NFL mRNA whereas normal SOD1 does not. Moreover, 14-3-3 and trace amine receptor (TAR) DNA-binding protein 43 (TDP-43) are two proteins incorporated in ALS intraneuronal aggregates that bind and destabilize NFL mRNA,<sup>40,41</sup> a phenomenon that could contribute to the aggregation of NFs in ALS.

## Mouse Models with Neurofilament Abnormalities

In the last two decades, the gene-targeting technique and transgenic mouse approaches have been used to investigate the contribution of cytoskeletal abnormalities in ALS pathogenesis (see *Table 2*). Evidence that NF disorganization *in vivo* can provoke neuronal death came from the observation that expression of a mutated NFL protein<sup>42</sup> or of an excess of peripherin<sup>36</sup> in transgenic mice induced the formation of ALS-like NF aggregates and selective degeneration of spinal motor neurons.

Abnormal NF accumulations have been detected in familial ALS cases due to *SOD1* mutations.<sup>43</sup> Similarly, transgenic mice expressing mutant SOD1 exhibit NF accumulations<sup>44,45</sup> and defective axonal transport of NFs in motor neurons.<sup>20,21</sup> To determine whether axonal NFs are involved in *SOD1*-mediated disease, mice expressing mutant SOD1 were mated with mice having low axonal NF content.<sup>46,47</sup> The results indicated that axonal NFs are not required for *SOD1*-mediated disease. Nonetheless, the absence of NFL in SOD1<sup>G85R</sup> caused depletion of axonal NFs and extension of lifespan by approximately 15%.<sup>46</sup> Surprisingly, overexpression of hNFH in SOD1<sup>G37R</sup> mice<sup>47</sup> and of mouse NFL or mouse NFH in SOD1<sup>G93A</sup> mice<sup>48</sup> also increased their lifespan by 65 and 15%, respectively. However, the mechanism of protection is still unclear. NF proteins contain multiple calcium binding sites.<sup>49</sup>

Therefore there is a possibility that perikaryal accumulations of NF proteins may act as calcium chelators to confer neuroprotection in a manner reminiscent of the calcium-binding protein calbindin-D28k when overexpressed in cultured motor neurons.<sup>50</sup> Nguyen et al.<sup>51</sup> also proposed that perikaryal accumulations of NFs in motor neurons may alleviate ALS pathogenesis by acting as a phosphorylation sink for cyclin-dependent kinase 5 (CDK5) dysregulation induced by mutant SOD1, thereby reducing the detrimental hyperphosphorylation of tau and other neuronal substrates. However, this hypothesis has been challenged because removal of NFM and NFH sidearms also led to a delay of disease in SOD1 mutant mice,<sup>52</sup> probably through enhancement of anterograde axonal transport.

Alternatively, such apparent discrepancies may reflect the existence of multiple neuroprotection mechanisms. It is noteworthy that hNFH overexpression conferred more robust protective effects than deletion of NF sidearms, with lifespan extension of six months compared with two months in the SOD1<sup>G37R</sup> mice.<sup>39,43</sup> The superior protective effects of hNFH overexpression could have resulted from depletion of axonal NF content together with phosphorylation sink of perikaryal NF accumulations. Finally, Ehlers et al.<sup>53</sup> have shown that NFs are involved in localisation of NMDA receptors in the neuronal plasma membrane by interacting with the NMDA NR1 sub-unit, suggesting that accumulation of NFs may interfere with glutamate receptor function. There is a report that NF-aggregate-bearing neurons exhibit increased intracellular calcium levels and enhanced cell death in response to NMDA-mediated excitotoxicity.<sup>54</sup>

## Conclusion

Accumulation of NFs in ALS was described for the first time several decades ago. Nevertheless, how the NF accumulations arise and the extent to which they contribute to disease pathogenesis is not fully elucidated. Lines of evidence suggest that perturbations of axonal transport or of IF protein stoichiometry can contribute to formation of intraneuronal IF inclusions, these two mechanisms not being mutually exclusive. Alterations in the stoichiometry of IF proteins can result from destabilization of IF mRNAs. Aberrant phosphorylation of NFs can also affect their axonal transport, but this phenomenon is poorly understood. A regulation of NF interactions to molecular motors is most probably involved in controlling NF movement. In order to develop a therapeutic approach able to restore a normal axonal transport of NFs in ALS, it will be important to identify which molecular motors and adaptor proteins are involved in this process. In mouse models, some types of NF accumulation correlate with motor neuron degeneration, whereas others seem to confer protection. The toxicity or benefit of NF accumulations could be due to differences in their localization (perikaryal versus axonal) and their ability to block fast axonal transport and sequester other vital organelles, such as mitochondria. ■



Rodolphe Perrot, PhD, is a Post-doctoral Fellow in the Department of Anatomy and Physiology at the Research Center at Laval University Hospital Center in Québec. His research focuses on investigating the pathogenic mechanisms associated with disorganization of the neuronal cytoskeleton. He obtained his PhD in molecular and cellular biology in 2006 at the Laboratory of Neurobiology and Transgenesis at Angers University Hospital in France.



Jean-Pierre Julien, PhD, is a Professor and holds a Canada Research Chair in the Department of Anatomy and Physiology at the Research Center at Laval University Hospital Center. Prior to this, he was a Professor at McGill University. His studies with genetically modified mice led to the first demonstration that disorganization of intermediate filaments may cause neurological disease. In 2000, Professor Julien received the Sheila Essey Award for research on

amyotrophic lateral sclerosis (ALS) from the American Academy of Neurology (AAN). He obtained his PhD from McGill University and carried out post-doctoral work at the National Institute for Medical Research (NIMR) in the UK.

1. Perrot R, Berges R, Bocquet A, et al., *Mol Neurobiol*, 2008;38:27–65.
2. Corbo M, Hays AP, *J Neuropathol Exp Neurol*, 1992;51:531–7.
3. Manetto V, Sternberger NH, Perry G, et al., *J Neuropathol Exp Neurol*, 1988;47:642–53.
4. Wong NK, He BP, Strong MJ, *J Neuropathol Exp Neurol*, 2000;59:972–82.
5. Niebroj-Dobosz I, Dziewulska D, Janik P, *Folia Neuropathol*, 2006;44:191–6.
6. Zetterberg H, Jacobsson J, Rosengren L, et al., *Eur J Neurol*, 2007;14:1329–33.
7. Figlewicz DA, Krizus A, Martinoli MG, et al., *Hum Mol Genet*, 1999;8:157–61.
8. Tomkins J, Usher P, Slade JY, et al., *Neuroreport*, 1998;9:3967–70.
9. Al-Chalabi A, Andersen PM, Nilsson P, et al., *Hum Mol Genet*, 2006;44:157–64.
10. Rooke K, Figlewicz DA, Han FY, et al., *Neurology*, 1996;46:789–90.
11. Vechio JD, Bruijn LI, Xu Z, et al., *Ann Neurol*, 1996;40:603–10.
12. Mersivanova IV, Perepelov AV, Polyakov AV, et al., *Am J Hum Genet*, 2000;67:37–46.
13. De Jonghe P, Mersivanova I, Nelis E, et al., *Ann Neurol*, 2001;49:245–9.
14. Georgiou DM, Zidar J, Korosec M, et al., *Neurogenetics*, 2002;4:93–6.
15. Yoshihara T, Yamamoto M, Hattori N, et al., *J Peripher Nerv Syst*, 2002;7:221–4.
16. Gros-Louis F, Lariviere R, Gowing G, et al., *J Biol Chem*, 2004;279:45951–6.
17. Chou SM, Wang HS, Taniguchi A, et al., *Mol Med*, 1998;4:324–32.
18. Crow JP, Ye YZ, Strong M, et al., *J Neurochem*, 1997;69:1945–53.
19. Strong MJ, Sopper MM, Crow JP, et al., *Biochem Biophys Res Commun*, 1998;248:157–64.
20. Williamson TL, Cleveland DW, *Nat Neurosci*, 1999;2:50–56.
21. Collard JF, Cote F, Julien JP, *Nature*, 1995;375:61–4.
22. Zhang B, Tu P, Abtahian F, et al., *J Cell Biol*, 1997;139:1307–15.
23. Ackerley S, Grierson AJ, Brownlees J, et al., *J Cell Biol*, 2000;150:165–76.
24. Manser C, Stevenson A, Banner S, et al., *FEBS Lett*, 2008;582:2303–8.
25. King AE, Dickson TC, Blizzard CA, et al., *Eur J Neurosci*, 2007;26:2151–9.
26. Kesavapany S, Patel V, Zheng YL, et al., *Mol Biol Cell*, 2007;18:3645–55.
27. Hafezparast M, Klocke R, Ruhrberg C, et al., *Science*, 2003;300:808–12.
28. LaMonte BH, Wallace KE, Holloway BA, et al., *Neuron*, 2002;34:715–27.
29. Xia CH, Roberts EA, Her LS, et al., *J Cell Biol*, 2003;161:55–66.
30. Motil J, Dubey M, Chan WK, et al., *Brain Res*, 2007;1164:125–31.
31. Teuling E, van Dis V, Wulf PS, et al., *Hum Mol Genet*, 2008;17:2849–62.
32. Xu Z, Cork LC, Griffin JW, et al., *Cell*, 1993;73:23–33.
33. Wong PC, Marszalek J, Crawford TO, et al., *J Cell Biol*, 1995;130:1413–22.
34. Cote F, Collard JF, Julien JP, *Cell*, 1993;73:35–46.
35. Meier J, Couillard-Despres S, Jacomy H, et al., *J Neuropathol Exp Neurol*, 1999;58:1099–1110.
36. Beaulieu JM, Nguyen MD, Julien JP, *J Cell Biol*, 1999;147:531–44.
37. Beaulieu JM, Jacomy H, Julien JP, *J Neurosci*, 2000;20:5321–8.
38. Millecamps S, Robertson J, Lariviere R, et al., *J Neurochem*, 2006;98:926–38.
39. Ge WW, Wen W, Strong W, et al., *J Biol Chem*, 2005;280:118–24.
40. Ge WW, Volkening K, Leystra-Lantz C, et al., *Mol Cell Neurosci*, 2007;34:80–87.
41. Strong MJ, Volkening K, Hammond R, et al., *Mol Cell Neurosci*, 2007;35:320–27.
42. Lee MK, Marszalek JR, Cleveland DW, *Neuron*, 1994;13:975–88.
43. Rouleau GA, Clark AW, Rooke K, et al., *Ann Neurol*, 1996;39:128–31.
44. Tu PH, Raju P, Robinson KA, et al., *Proc Natl Acad Sci U S A*, 1996;93:3155–60.
45. Borchelt DR, Wong PC, Becher MW, et al., *Neurobiol Dis*, 1998;5:27–35.
46. Williamson TL, Bruijn LI, Zhu Q, et al., *Proc Natl Acad Sci U S A*, 1998;95:9631–6.
47. Couillard-Despres S, Zhu Q, Wong PC, et al., *Proc Natl Acad Sci U S A*, 1998;95:9626–30.
48. Kong J, Xu Z, *Neurosci Lett*, 2000;281:72–4.
49. Lefebvre S, Mushynski WE, *Biochem Biophys Res Commun*, 1987;145:1006–11.
50. Roy J, Minotti S, Dong L, et al., *J Neurosci*, 1998;18:9673–84.
51. Nguyen MD, Lariviere RC, Julien JP, *Neuron*, 2001;30:135–47.
52. Lobsiger CS, Garcia ML, Ward CM, et al., *Proc Natl Acad Sci U S A*, 2005;102:10351–6.
53. Ehlers MD, Fung ET, O'Brien RJ, et al., *J Neurosci*, 1998;18:720–30.
54. Sanelli T, Strong MJ, *Free Radic Biol Med*, 2007;42:143–51.
55. Graham DG, *Curr Opin Neurol*, 1999;12:733–7.
56. Chou SM, Hartmann HA, *Acta Neuropathol*, 1965;4:590–603.
57. Griffin JW, Hoffman PN, Clark AW, et al., *Science*, 1978;202:633–5.
58. Griffin JW, Parhad I, Gold B, et al., *Neurotoxicology*, 1985;6:43–53.
59. Eyer J, McLean WG, Lettieri JF, *J Neurochem*, 1989;52:1759–65.
60. Zhu Q, Lindenbaum M, Levasseur F, et al., *J Cell Biol*, 1998;143:183–93.
61. Gold BG, Griffin JW, Price DL, *J Neurosci*, 1985;5:1755–68.
62. Tanii H, Hayashi M, Hashimoto K, *Arch Toxicol*, 1988;62:70–75.
63. Endo H, Kittur S, Sabri MI, *Neurochem Res*, 1994;19:815–20.
64. Reagan KE, Wilmarth KR, Friedman M, et al., *Neurochem Int*, 1994;25:133–43.
65. Kadota T, Kadota K, *J Toxicol Sci*, 1978;3:57–67.
66. Nixon RA, Clarke JF, Logvinenko KB, et al., *J Neurochem*, 1990;55:1950–59.
67. Shea TB, Balikian P, Beermann ML, *FEBS Lett*, 1992;307:195–8.
68. Shea TB, Wheeler E, Jung C, *Mol Chem Neuropathol*, 1997;32:17–39.
69. Vahidnia A, Romijn F, Tiller M, et al., *Hum Exp Toxicol*, 2006;25:667–74.
70. DeFuria J, Shea TB, *Brain Res*, 2007;1181:74–82.
71. Valentine WM, Amarnath V, Graham DG, et al., *Toxicol Appl Pharmacol*, 1997;142:95–105.
72. Valentine WM, Graham DG, Anthony DC, *Toxicol Appl Pharmacol*, 1993;121:71–7.
73. Lurie DI, Brooks DM, Gray LC, *Neurotoxicology*, 2006;27:108–17.
74. Jones LG, Prins J, Park S, et al., *J Comp Neurol*, 2008;506:1003–17.
75. Jensen KF, Lapadula DM, Anderson JK, et al., *J Neurosci Res*, 1992;33:455–60.
76. Zhao XL, Zhang TL, Zhang CL, et al., *Toxicology*, 2006;223:127–35.
77. Gama Sosa MA, Friedrich VL Jr., DeGasperi R, et al., *Exp Neurol*, 2003;184:408–19.
78. Zhu Q, Couillard-Despres S, Julien JP, *Exp Neurol*, 1997;148:299–316.
79. Kriz J, Zhu Q, Julien JP, et al., *Brain Res*, 2000;885:32–44.
80. Yuan A, Rao MV, Kumar A, et al., *J Neurosci*, 2003;23:9452–8.
81. Dubois M, Strazielle C, Julien JP, et al., *J Neurosci Res*, 2005;80:751–8.
82. Xu Z, Marszalek JR, Lee MK, et al., *J Cell Biol*, 1996;133:1061–9.
83. Beaulieu JM, Julien JP, *J Neurochem*, 2003;85:248–56.
84. Ishihara T, Hong M, Zhang B, et al., *Neuron*, 1999;24:751–62.
85. Kieran D, Hafezparast M, Bohnert S, et al., *J Cell Biol*, 2005;169:561–7.
86. Laird FM, Farah MH, Ackerley S, et al., *J Neurosci*, 2008;28:1997–2005.