

Neuroinflammatory Cytokines – The Common Thread in Alzheimer Pathogenesis

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Abstract

This article discusses the potential role of the cytokine cycle and its corollary as drivers of the relentless progression of Alzheimer's neuropathologies, whether they are the result of gene mutations, gene polymorphisms and/or environmental and comorbid conditions. Based on the discovery of cytokine overexpression as an accompaniment to the dementia-related glial activation, the cytokine hypothesis was proposed. This states that in response to the negative impact on neurons of known and unknown risk factors – which include genetic inheritance, comorbid and environmental factors – microglia and astrocytes become activated and produce excess amounts of the immune-modulating cytokine interleukin-1 (IL-1) and the neurotogenic cytokine S100B, respectively. Finding that these glial events occur in foetuses and neonates with Down's syndrome provided the first evidence that productive immune responses by activated glia precede rather than follow overt AD-related pathology. This finding can be added to the demonstration of IL-1 induction of amyloid- β (A β) precursor protein and astrocyte activation with excess production of neurotogenic factor S100B. This combination suggests that IL-1 and S100B overexpression would favour the A β production and dystrophic neurite growth necessary for laying down neuritic A β plaques. This, together with demonstration of IL-1 induction of excessive production of the precursors of other features common in AD prompted a corollary to the cytokine hypothesis. The corollary states that regardless of the primary cause of the neuronal insult, the result will be chronic glial activation, which in turn will result in further neuronal injury, still more glial activation with excess cytokine expression and so on. This article discusses known causes, genetic and environmental risk factors, and comorbid conditions and the potential contribution of glial activation with excessive cytokine expression to each.

Keywords

Alzheimer's disease, amyloid β , apolipoprotein E, β -amyloid precursor protein, cytokines, Down's syndrome, excitotoxicity, glutamate, interleukin-1, S100B, tumour necrosis factor alpha

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Alzheimer's disease (AD) is the epitome of a progressive degenerative disease of the brain and culminates in dementia characterised in the beginning stages as a failure to retain new information for appropriate retrieval. As the underlying disease progresses, the patient experiences more profound memory loss and retrieval dysfunctions. These include not only the loss of skills related to activities of daily living, but may also include personality changes associated with inappropriate or psychotic behaviours. These changes in cognitive, behavioural and social functions are now known to be preceded by neuropathological changes in regions of the brain involved in registering, storing and recalling new information in appropriate circumstances and for the appropriate duration. The most striking and diagnostic neuropathological change in the Alzheimer's brain is the presence of amyloid- β (A β) plaques lying within the neuronal layers in cortical regions that are important in acquiring, storing and retrieving information. These regions include

temporal lobe structures, such as the hippocampus, as well as frontal and parietal regions. It is the number of A β plaques that contain dystrophic neuronal processes (A β neuritic plaques) in affected brain regions that provide the neuropathological confirmation of the clinical diagnosis of AD. Many neurons within these plaque-laden cortical layers contain within their cytoplasm fibrillary tangles composed of hyperphosphorylated tau coiled into paired helical filaments, the so-called neurofibrillary tangles.

A progression of these neuropathological events seems apparent. For example, the plethora of A β plaques at end stages – from several to many – and an inclusion of neurofibrillary tangles – in a few to numerous neurons – strongly suggests that these neuropathological changes are cumulative. Similarly, progression of plaques from a non-neuritic, diffuse appearance to one of compact deposits is suggested by the appearance of different A β plaque types. These

include diffuse A β deposits, diffuse neuritic A β plaques, dense-core neuritic A β plaques and dense core non-neuritic A β plaques within a single tissue section.

Neurofibrillary progression is suggested by the formation of 'threads' of paired helical filaments scattered within neuronal cytoplasm, along with other cells in which fully-formed tau tangles push aside the nucleus and acellular tangles that appear to have remained after the death and dissolution of a neuron.

Such progression suggests the presence of driver(s) of these events, which, even without repetition of the causative event(s) or agent(s), result in self-propagation of the neurodegenerative events that culminate in increasing densities of these neuropathological changes. This article discusses the common themes in neuropathogenesis:

- potential causative factors (gene mutations and gene duplication);
- risk-conferring factors (gene polymorphisms, environmental and comorbid conditions and ageing); and
- omnipresent glial activation and excessive expression of putative drivers (these have been found to be factors that promote furtherance of neurodegenerative events).

Causative Factors

Gene Mutations – *APP* and *PSEN1* and *PSEN2*

A small number of families worldwide have a high percentage of individuals who develop AD at an early age, usually well before 65 years of age, suggesting autosomal-dominant inheritance. Glenner's discovery that the so-called senile plaques in the brain of AD patients are largely composed of A β highlighted the importance of this peptide in AD pathogenesis.^{1,2} This discovery was quickly followed by others demonstrating the structure and size of the A β peptide necessary for fibril formation^{3,4} and mapping of the amyloid precursor protein (*APP*) gene for the β -amyloid precursor protein (β APP) to chromosome (Chr) 21.⁵⁻⁸

The known occurrence of families with autosomal-dominantly inherited AD led to studies linking mutations in the β APP coding region to familial AD (FAD).^{9,10} Although additional family-linked *APP* gene mutations were identified in subsequent searches, they did not account for all the known familial cases. Studies of such families revealed specific mutations in two other gene sequences that are causative for AD: presenilins (*PSEN*) 1 and 2. These genes are located on Chr 1 (*PSEN2*) and Chr 14 (*PSEN1*).¹¹⁻¹⁴ Inheritance of any one of these autosomal-dominant, highly penetrant missense mutations in the *APP* or *PSEN* genes causes the development of early-onset AD. More recently, early-onset cases have been attributed to promoter mutations that merely elevate the expression of β APP.¹⁵

Taken together, mutations in the *APP*, *PSEN1*, and *PSEN2* genes account for the vast majority of familial cases.¹⁶ Of the three genes, the 173 recognised mutations in *PSEN1* are responsible for most cases.¹⁷ Overproduction of longer forms of A β is the common thread between these mutations¹⁷ and families with them exhibit the entire array of neuropathological changes characteristic of AD at an accelerated pace, resulting in early onset. It is notable that increased expression of β APP is characteristic of a number of lifestyle and environmental factors associated with increased risk for later development of AD. Moreover, lifestyle risks and genetic factors are likely additive and, thus, may accelerate onset.¹⁸

Insights into the mechanistic relationships between these mutations and neuropathology have been afforded through transgenesis of mice. These have had either wild-type¹⁹ or mutated human β APP sequences,^{20,21} mutated *PSEN1*,²²⁻²⁵ mutated *PSEN1* plus mutated β APP^{26,27} or these plus mutated human tau.²⁸ Mouse models have produced evidence of glial activation,^{29,30} as well as specific receptors and related signaling molecules³¹⁻³³ or specific cytokines and other inflammatory factors³⁴⁻³⁶ related to Alzheimer's pathogenesis. Such models have also been used to examine broader phenomena and potential interventions, including the protection afforded by exercise³⁷ and the benefits of non-steroidal anti-inflammatory drugs (NSAIDs).³⁸⁻⁴³ Taken together, these findings and the many others in which transgenic mice were used have added greatly to the understanding of molecular biological mechanisms of AD pathogenesis. They have also shown mechanisms, cytokines and other factors that may act as drivers of such mechanisms.

Gene Duplications – Down's Syndrome

A link between Chr-21 genes and Down's syndrome (DS; trisomy 21) was suggested by the finding of Alzheimer's-like clinical and neuropathological changes at middle age in individuals with DS.^{44,45} This suggestion was further supported by mapping of the *APP* gene to chromosome 21.^{5,6,10,46,47} It is conceivable that any of the other Chr-21 genes could contribute to AD pathology in conventional cases of trisomy, but DS resulting from a partial rearrangement of Chr-21 sequences excluding the *APP* gene was not associated with precocious AD pathology.⁴⁸ More recently, duplications of the *APP* locus alone have been identified in French, Dutch and Japanese pedigrees of FAD.^{49,50} None of the affected individuals has DS, parsing AD symptomatology with the *APP* gene itself. Duplication of the *APP* gene in either DS or FAD elevates its messenger RNA (mRNA) levels by about 50 %,⁵¹ which is consistent with faithful transcription of the 1.5-fold gene number. The steady-state levels of β APP protein are greater than expected, however, from the 1.5-fold gene load in trisomy.^{5,52} These increases do not occur until advanced age in either DS⁵³ or mouse models thereof,⁵⁴ the increases may even be brain-specific.⁵⁵

These circumstances suggest that the expected increase owing to gene loading is augmented by factors that act to elevate β APP protein levels, perhaps including activated glia and the cytokines they overexpress in the mature brain.^{56,57} This idea, together with previous studies suggesting a role for glia in plaque formation,^{58,59} is supported by a series of investigations demonstrating the potential of these cytokines to act as generators and promoters of the neuropathological sequelae of AD, i.e.:

- A β plaques;⁶⁰⁻⁶²
- neurofibrillary tangles;⁶³⁻⁶⁶
- dysregulation of neurotransmitters;⁶⁷
- growth factor levels;⁶⁸ and
- DNA damage and neuron loss.⁶⁹⁻⁷²

A perhaps more accessible link between AD and DS neuropathologies is afforded by a recently discovered connection between lens pathology in DS and AD, i.e. accumulation of A β in the lens and accompanying accumulation in the brain.⁷³ A great deal of work has been accomplished using mouse models of DS⁷⁴ as well as DS fibroblast cell lines.^{54,75-78} In particular, work has demonstrated that β APP fragments, but not A β , are responsible for disruptions in the endosomal dysfunction associated with failure to clear unwanted proteins.⁷⁸

The Common Thread – Glial Activation and Cytokine Expression

Familial Alzheimer's Disease

In a neuropathological analysis of cerebellar tissue from FAD (APP Val717 → Ile: V717F), glial activation was consistently and markedly elevated compared to sporadic AD (SAD). This was correlated with a 25 % decline in Purkinje cell numbers.⁷⁹ By contrast, A β plaque density – whether extracellular, vascular or perivascular – did not differ between FAD and SAD tissues.

Associations have already been made in AD between increased glial activation and excessive cytokine expression and cell death,^{70,71,80} and similar relationships have been observed in mouse models.⁸¹ Both human and mouse models, as well as the relationship between cytokines and neurodegeneration, are discussed in a recent review.⁸²

Down's Syndrome

DS has been suggested as a human model for studying the progression and potential drivers of AD pathogenesis.⁸³ This is because:

- the extra copy of Chr 21 in DS is present in the zygote so that all the players, including the 1.5-fold gene loading and its downstream consequences, are in place at conception; and
- the innate and, to date unstoppable, outcome is AD.

Microglia and astrocytes had previously been found expressing excessive amounts of the systemic immune response-generating cytokine interleukin-1 (IL-1) and astrocyte-derived neurotogenic cytokine S100B, respectively, in the AD brain. Considering the relationships between DS and AD, Griffin et al. set out to assess elements of an innate immune response in the DS brain. They wanted to explore the possibility that changes that are nearly universal in the DS brain would render it a suitable system for studying drivers of AD pathogenesis.

Compared with the brains of non-DS individuals of similar ages, IL-1 and S100B were highly overexpressed in activated glia, even in the brains of fetuses, neonates and children with DS.⁵⁶ This age distribution is important; neither A β plaques nor neurofibrillary tangles were noted in these brains, nor have these AD neuropathological anomalies been reported in brains from DS individuals of these ages.^{84,85} Thus, several studies underscore the importance of these cytokines and precocious development of A β plaques and neurofibrillary tangles in DS. These show:

- the increase in IL-1 and S100B expression in DS prior to the detection of either A β plaques or neurofibrillary tangles;^{56,57}
- the capacity of both IL-1⁸⁶ and S100B to elevate neuronal β APP;⁸⁷
- the induction of tau phosphorylation through IL-1-induced activation of mitogen-activated protein kinase p-38;⁴⁵ and
- the relation of overexpression of IL-1 and S100B to both A β plaque^{62,88,89} and neurofibrillary tangle^{63,64} development.^{57,83}

In addition to these DS-related changes that are accompanied by dramatically increased levels of β APP,⁹⁰ the resulting overexpression of the β APP cleavage fragment β CTF causes endosomal dysfunction.⁷⁸ Griffin et al. propose that this β CTF expression-dependent increase in endosomal dysfunction is indirectly related to the upregulation of IL-1 by excessive DS-related increases in β APP and the resulting release of another β APP cleavage fragments secreted (s)APP. Such sAPP induces glial activation and excessive production and release of IL-191 and

bCTF, which results in endosomal dysfunction and leads to the build-up of unwanted proteins. This induces glial activation and excessive production and release of IL-1⁹¹ and β CTF, which results in endosomal dysfunction and leads to the buildup of unwanted proteins.⁹²

These events may create another opportunity for progression through positive feed-forward. The APP intracellular domain, produced by γ -secretase cleavage of β CTF, is capable of promoting AD-related symptoms – including tau pathology and inhibition of neurogenesis – in a manner sensitive to NSAIDs.⁹³ This may be related to the expanding evidence for an inhibition of neurogenesis by IL-1.^{94,95}

Animal Models

The creation of mouse models of FAD through transgenesis with mutations or deletions of *APP*, *PSEN1* and *PSEN2* have greatly aided in the understanding of AD pathogenesis.

Glial activation is a prominent feature in these models from the first FAD-mutant APP (V717F) transgenic mouse model.²⁰ In more recent studies, astrocyte activation in V717F transgenic mice was shown to be accompanied by microglial activation months before the appearance of A β plaques in these animals.

Compared with littermate-controls (non-transgenic mice), V717F mice had elevated numbers of activated astrocytes overexpressing S100B as early as two months of age, while the first A β plaques were not found until eight months.⁹⁶ The tissue levels of S100B mRNA and protein were elevated at each month from the second to eighth and from that time were correlated with A β load in these mice.⁹⁶ In addition, the expression of the IL-1 β -converting enzyme is greatly increased in these mice, again before plaque formation, and was closely correlated with neuronal DNA damage and cell loss. This is consistent with the idea that increases in active IL-1 β contribute to A β plaque formation and to neuronal degeneration in mouse models of AD and by analogy to AD.⁷⁰ Accompanying this astrocyte activation in V717F transgenic mice, there is microglial activation with overexpression of IL-1 and other inflammation parameters, again prior to A β plaque deposition.⁹⁷

Crossing another APP-transgenic mouse (Tg2576) with transgenic mice overexpressing S100B (TghuS100B) exaggerated the glial activation and increased expression of S100B above the level in the TghuS100B mice. It also increased expression of IL-1 β , IL-6 and tumour necrosis factor alpha (TNF- α) above the expression level in Tg2576 mice.³⁴ The increase in glial activation and overexpression of these cytokines was an early phenomenon that foreshadowed an increase in plaque density and A β levels.³⁴

A study of a mouse model in which PS-1 and PS-2 functions were eliminated (*PSEN1* null mutation with conditional knockout of *PSEN2*) found:⁹⁸

- substantial glial activation;
- neuronal loss in the cortex and hippocampus;
- cortical and hippocampal atrophy; and
- tau hyperphosphorylation with tangle-like intraneuronal inclusions.

All of this occurred with no additional production of A β . The activation of glia and the accompanying increase in cytokines, such as S100B

and IL-1, may contribute to this neuronal dysfunction and loss, as demonstrated by Mori et al.³⁴ and Sheng et al.⁷⁰

Growing evidence has suggested beneficial roles for microglial activation in transgenic mouse models of A β deposition. For instance, plaque burdens are increased by deficiencies in the chemokine receptor CCR2⁹⁹ or in the complement system.⁹⁹ Indeed, A β deposition occurs efficiently in the near absence of microglia.¹⁰⁰ Nevertheless, considerable evidence indicates that activated microglia and their products are detrimental in many experimental models and natural disease states. It is likely that microglia could contribute to A β removal while simultaneously producing agents that are detrimental to neurological structure/function and conducive to progression of pathology. As evidence, genetic ablation of the microglial fractalkine receptor CX₃CR1 prevents neuronal loss in an APP-transgenic line: 20305648.³⁷

Gene Polymorphisms as Risk Factors for Development of Alzheimer's Disease

Genetic anomalies, such as the gene mutations and duplication discussed above, account for very few of the many sporadic AD cases. Inheritance of variants in specific sequences, i.e. polymorphisms, of a relatively small number of genes may contribute to these sporadic cases. Several such polymorphisms have been implicated in modifying the risk for development of AD.

In this article, the discussion includes the most important polymorphism, the ϵ 4 variant of the apolipoprotein E (*APOE*) gene, which is moderately penetrant and confers high risk for development of AD. For the purpose of establishing a link between neuroinflammatory glial activation and excessive expression of proinflammatory cytokines, the genetic links to just two of these cytokines, TNF α and IL-1, are reviewed here. This exclusivity leaves further discussion of other important factors involved in AD pathogenesis to another review. These include components of the complement system and many other factors^{101–103} that doubtless contribute to such pathogenesis and are deserving of a more sharply focused discussion.

Apolipoprotein E Gene

The gene polymorphism that is most important in AD risk is a single nucleotide substitution in the coding region of the *APOE* for apolipoprotein E (ApoE) on Chr 19, creating what is termed the ϵ 4 allele. The resulting gene product – ApoE4 – has an arginine–cysteine substitution at position 112 of its amino acid sequence. This alters the interaction of the protein with its receptor and increases AD risk.^{104,105} Individuals with two copies of the ϵ 4 allele of *APOE* have a 50–90 % chance of developing AD by the age of 85 and those with one copy have about a 45 % chance.

The mechanisms underlying the genetic influence of *APOE* ϵ 4 on AD risk are unknown, but hypotheses include an interaction with A β as plaques develop.¹⁰⁶ Inheritance of *APOE* ϵ 4 is associated with increased A β plaque density¹⁰⁷ as well as neuronal pathologies.¹⁰⁸ In addition to this, ApoE is important for the molecular trafficking¹⁰⁹ and induction of neuronal expression of β APP, perhaps in particular with regard to specific genotypes of *APOE*. This is because the *APOE* ϵ 3 variant is more effective at β APP induction than *APOE* ϵ 4.¹¹⁰

A good deal of evidence indicates an impact of *APOE* genotype on inflammatory sequelae. *APOE* ϵ 4 was found to produce a

dose-dependent increase in microglial activation and this effect was independent of A β deposition.¹¹¹ The protective effect of NSAID use is much stronger in *APOE* ϵ 4 carriers;^{112–116} indeed, some of these studies have restricted a significant NSAID effect to ϵ 4 carriers alone. This may explain the failure of several prospective trials with NSAIDs.^{117–119} The only trial showing a benefit was the only one powered to detect an effect of *APOE* genotype.¹²⁰ Mechanisms to explain the interaction between *APOE* genotype and inflammatory events are suggested by studies showing that microglia are more robustly activated in the presence of human ApoE4.^{91,121–123} This may involve differential inhibition of inflammatory agents, such as sAPP by ApoE3 and ApoE4.⁹¹

Reports from the first phase of the multicenter AD Neuroimaging Initiative (ADNI) are making major contributions to knowledge concerning correlations between neuroimaging and cerebrospinal fluid (CSF) analytes and AD progression. They are also helping discern the role of environmental, comorbid conditions and specific gene polymorphisms in increased risk or perhaps precocious development of AD neuropathological changes. For instance in a new report from ADNI, genotypes, including *APOE* and two other 'promising genes' contactin 5 (*CNTN5*) and bridging integrator 1 (*BIN1*), were associated with a number of neuroimaging parameters.¹²⁴ More specifically, the inheritance of *APOE* ϵ 4 and a maternal history of the disease were linked to hippocampal atrophy.¹²⁵

With regard to comorbid conditions, obesity is linked to lower brain volumes in AD and in mild cognitive impairment (MCI), a condition likely to be prodromal AD.¹²⁶ Importantly, there were indications of increased plaque burden by 3-D imaging with the 'Pittsburgh compound' PiB correlate with hippocampal atrophy¹²⁷ as well as with progression from MCI to AD.¹²⁸ Similar correlations hold with CSF measurements of A β and tau. Worldwide ADNI efforts will aid in establishing the progression from early stages, i.e. MCI to AD. Such efforts will then be useful in defining the mechanisms involved in driving AD neuropathology as well as developing and analysing rational strategies for therapeutic interventions.¹²⁹

Tumour Necrosis Factor Alpha

In a study to identify polymorphisms in *TNF*, the gene encoding the proinflammatory cytokine TNF α , a polymorphism at -850 C/T in *TNF* was found to increase the risk for AD.¹³⁰ Specific polymorphisms in promoter regions of inflammatory cytokine genes, such as TNF α and IL-6, are associated with late-onset sporadic AD.¹³¹ For instance, heterozygosity for a polymorphism at -1082 of the IL-10 gene increased the odds ratio of AD two-fold. When combined with a polymorphism at -308 in *TNF*, the odds ratio was increased 6.5 times.

Interleukin-1

Both *IL1A* and *IL1B*, which encode proteins IL-1 α and IL-1 β , map to Chr 2. Polymorphisms in each of these genes have been associated with AD. Many, but not all, case-control studies of individuals who died with AD have shown that polymorphisms in either of these IL-1 genes are associated with increased odds for AD. The principal risk-conferring polymorphism in *IL1A* is located in the gene promoter region (-889 C/T), while that in *IL1B* is located either in the promoter region (-511 C/T) or in the coding region (+3954). In a number of studies, inheritance of a polymorphism at either -889 of *IL1A* or +3954 of *IL1B* was shown to increase the odds of AD by

two- to four-fold.^{132–137} Importantly, in another of study of living AD patients, a -889 polymorphism in *IL1A* was associated not only with increased an risk for AD (4.5 times) but also with earlier disease onset (seven to nine years earlier).¹³⁴ Although the number of studies supporting such an association is increasing, a few studies have failed to substantiate this.^{138,139} In one study, *IL1B* at -511 increased the odds for AD but the *IL1A* -889 polymorphism did not.¹⁴⁰ A strong association was shown between AD and *IL1B* +3954, but not -511 in another study.¹⁴¹ Other studies have found no relation between *IL1B*-511 and AD.^{142,143} Meta-analyses have supported an association for at least the -889 polymorphism in the *IL1A* gene.^{144–146} In the large Honolulu-Asia Aging Study, the authors concluded that certain loci within the IL-1 gene cluster were associated with AD.¹⁴⁷

There are likely multiple reasons for these conflicting reports of the link between IL-1 gene polymorphisms and AD risk. These might include the disparate population groups studied, sample size and contributions by other risk-modifying genes. Importantly toward this point, in an extremely rigorous analysis of AD-related polymorphisms,¹⁴⁸ a polymorphism at -889 in *IL1A* was suggested as one that approached significance. It may have failed to show linkage only owing to deficiencies in replication and the spectrum of populations studied. In a recent study of the importance of polymorphisms in cytokine genes, specific polymorphisms in both *IL1A* and *IL1B* were positively associated with AD. Polymorphisms in the genes for anti-inflammatory factors, such as IL-4 and IL-10, were strongly associated with a reduced risk for AD.¹⁴⁹

As with all risk-conferring factors – whether related to inheritance or environmental events – downstream outcomes may at least be additive, or at worst multiplied. The multigene and additive nature of quantitative-trait loci is in contrast to high-penetrance single-gene mutations, which cause rather than alter risk for disease. A case in point with regard to the inheritance of specific gene combinations, or haplotypes, and the additive nature of risks on outcomes is the dramatic increase in risk with inheritance of both *IL1A* and *IL1B* polymorphisms. Inheritance of the polymorphisms in both *IL1A* and *IL1B* is associated with a much greater risk (~10 times) of developing AD.^{132,133,150,151}

Age-related Alzheimer's Disease Neuropathological Changes Advancing Age

Ageing provides the backdrop for the progression of degenerative events in general, including the development of the clinical symptoms and neuropathological changes characteristic of AD. Only with ageing do the cognitive and functional declines, as well as the elevated densities of neuritic A β plaques in neuronal layers and neurofibrillary tangles within neurons, manifest themselves. The importance of ageing in neuropathogenesis is readily apparent. Even individuals who have causative factors – specific genetic mutations or DS, both of which provide virtual assurance of development of AD – reach adulthood or even middle-to-late age before overt signs appear. In addition to this, with advancing age cognitively normal individuals often have an abundance of neurofibrillary tangles and neuritic A β plaques that may approach densities consistent with a neuropathological diagnosis of AD (for a review, see Mrazek and Griffin¹⁵²). As it is uncommon for the densities of these anomalies to reach those necessary for a definitive diagnosis of AD, Mackenzie and colleagues¹⁵³ suggest that cognitive function is related to plaque

density and that ageing *per se* is not synonymous with A β deposition and cognitive failure.

Until recently, little could be discerned regarding the relevance of age-related accumulations of A β plaques and neurofibrillary tangles, especially as to whether they were portents of later development of AD. One study used *in vivo* A β labeling together with functional magnetic resonance imaging to test cognitively intact elderly individuals. Sperling and her colleagues found a relationship between A β deposit density and changes in a specific memory mode previously noted in AD patients.¹⁵⁴ These findings suggest that events during supposedly normal ageing may presage memory loss and neural dysfunctions related to learning and memory. In this way, they may contribute to later development of AD, with greater memory dysfunction and neuropathological change. The changes highlight the importance of defining seminal events and triggers of the progressive acquisition of such A β deposits. This will help toward the development of rational strategies to halt or impede such acquisition. In this way it may be possible to prevent or delay consequent neural dysfunction and dementia.

Some cognitively intact individuals have large numbers of non-neuritic A β deposits, but interestingly these deposits are not associated with activated microglia. This suggests that microglial activation contributes to the progression of A β deposits into the more mature neuritic A β plaques diagnostic of AD.¹⁵⁵ Even in individuals who are cognitively normal and who have few A β deposits or plaques, the density and activation state of astrocytes and microglia increase with advancing age. The levels of S100B expression gradually increase,^{57,156} while the levels of IL-1 remain stable until the sixth decade.¹⁵⁷ The latter finding suggests that in normal ageing, IL-1 levels are maintained at lower levels in order to delay neurodegenerative consequences.

Environmental Events that Contribute to Precocious (Age-inappropriate) Development of Alzheimer's Disease Neuropathological Changes

In addition to genetic mutation, gene duplication, gene polymorphisms and the wear and tear of time with ageing, a number of lifestyle or environmental risk factors have been associated with the precocious development of AD. Given the fact that epilepsy is often one of the consequences of head injury, it is likely that these problems exacerbate and/or hasten the precocious development of neuropathological changes of AD.

Indeed, this is the case when either epilepsy or head injury is assessed with regard to the influence of inheritance of an APOE ϵ 4 allele; such inheritance adds to the risk for a poor outcome in both epilepsy and head injury.^{158,159} Conditions associated with an increased risk for AD are as wide ranging as:

- heart disease;¹⁶⁰
- AIDS;^{161,162}
- obesity and diabetes;^{126,163–168} and
- hypertension and heart disease.^{165,168–170}

As with head injury and epilepsy, the risks conferred are increased in carriers of one or more APOE ϵ 4 alleles.^{164,170,171} In addition, head injury, epilepsy and AIDS include early overexpression of β APP and increases in neuroinflammatory cytokines, such as IL-1 and S100B.^{82,172–177} In several of the conditions mentioned above, glial activation has been

associated with neuropathological changes similar to those in AD; glial activation is sometimes present long before frank neurodegeneration is noted. Conditions involving glial activation include:

- DS;^{56,57}
- head injury;^{172,175}
- epilepsy;^{176,177}
- obesity and diabetes;^{126,166,167} and
- AIDS.¹⁷⁸

Summary

The pervasive appearance of glial activation and proinflammatory cytokine overexpression prior to and accompanying the neuropathological changes of AD suggests their importance in its development. The known capacity of at least one of these cytokines – IL-1 – to induce expression of other cytokines and the hallmarks of AD neuropathology highlights cytokine importance in Alzheimer's pathogenesis.

Discovery of proinflammatory activity exhibited by sAPP provided the missing link between the two universal responses of neural systems to injury: increased neuronal expression of APP and glial activation with excess cytokine expression. Namely, injury-induced overproduction of sAPP provides for a progressive feed-forward cycle whereby the resultant microglial activation and release of IL-1 leads to further β APP elevation.

Knowledge of the mechanisms involved in this link has provided a framework for understanding the way in which neuronal-glial interactions culminate in AD. Moreover, this also explains, at least in part, why neuronal trauma sets a cycle in motion. Neuronal trauma can be induced by untoward genetic inheritance, direct injury, comorbid conditions or as-yet unknown factors. The self-propagating cycle, properly called the cytokine cycle, promotes chronic production of glial cytokines, precursors of A β plaques, neurofibrillary tangles, Lewy bodies, the buildup of unwanted proteins and cognitive decline. ■

1. Glenner GG, Wong CW, Quaranta V, et al., The amyloid deposits in Alzheimer's disease: their nature and pathogenesis, *Appl Pathol*, 1984;2:357–69.
2. Glenner GG, Wong CW, Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein, *Biochem Biophys Res Commun*, 1984;120:885–90.
3. Gorevic PD, Castano EM, Sarma R, et al., Ten to fourteen residue peptides of Alzheimer's disease protein are sufficient for amyloid fibril formation and its characteristic x-ray diffraction pattern, *Biochem Biophys Res Commun*, 1987;147:854–62.
4. Gorevic PD, Goñi F, Pons-Estel B, et al., Isolation and partial characterization of neurofibrillary tangles and amyloid plaque core in Alzheimer's disease: immunohistological studies, *J Neuropathol Exp Neurol*, 1986;45:647–64.
5. Tanzi RE, Gusella JF, Watkins PC, et al., Amyloid beta protein gene: cDNA, mRNA distribution, and genetic linkage near the Alzheimer locus, *Science*, 1987;235:880–4.
6. Watkins PC, Tanzi RE, Cheng SV, et al., Molecular genetics of human chromosome 21, *J Med Genet*, 1987;24:257–70.
7. Goldgaber D, Lerman MI, McBride OW, et al., Characterization and chromosomal localization of a cDNA encoding brain amyloid of Alzheimer's disease, *Science*, 1987;235:877–80.
8. Korenberg JR, Pulst SM, Neve RL, et al., The Alzheimer amyloid precursor protein maps to human chromosome 21 bands q21.105-q21.05, *Genomics*, 1989;9:124–7.
9. St George-Hyslop PH, Tanzi RE, Polinsky RJ, et al., The genetic defect causing familial Alzheimer's disease maps on chromosome 21, *Science*, 1987;235:885–90.
10. Cheng SV, Nadeau JH, Tanzi RE, et al., Comparative mapping of DNA markers from the familial Alzheimer disease and Down syndrome regions of human chromosome 21 to mouse chromosomes 16 and 17, *Proc Natl Acad Sci U S A*, 1988;85:6032–6.
11. Cruts M, Hendriks L, Van Broeckhoven C, The presenilin genes: a new gene family involved in Alzheimer disease pathology, *Hum Mol Genet*, 1996;5:1449–55.
12. Kovacs DM, Fausett HJ, Page KJ, et al., Alzheimer-associated presenilins 1 and 2: neuronal expression in brain and localization to intracellular membranes in mammalian cells, *Nat Med*, 1996;2:224–9.
13. Sherrington R, Rogaeve EI, Liang Y, et al., Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease, *Nature*, 1995;375:754–60.
14. Levy-Lahad E, Wasco W, Poorkaj P, et al., Candidate gene for the chromosome 1 familial Alzheimer's disease locus, *Science*, 1995;269:973–7.
15. Theuns J, Brouwers N, Engelborghs S, et al., Promoter mutations that increase amyloid precursor-protein expression are associated with Alzheimer disease, *Am J Hum Genet*, 2006;78:936–46.
16. Hutton M, Busfield F, Wrang M, et al., Complete analysis of the presenilin 1 gene in early onset Alzheimer's disease, *Neuroreport*, 1996;7:801–5.
17. Shepherd C, McCann H, Halliday GM, Variations in the neuropathology of familial Alzheimer's disease, *Acta Neuropathol*, 2009;118:37–52.
18. Whittington RA, Papon MA, Chouinard F, et al., Hypothermia and Alzheimer's disease neuropathogenic pathways, *Curr Alzheimer Res*, 2010;7:17–25.
19. Quon D, Wang Y, Catalano R, et al., Formation of beta-amyloid protein deposits in brains of transgenic mice, *Nature*, 1991;352:239–41.
20. Games D, Adams D, Alessandrini R, et al., Alzheimer-type neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein, *Nature*, 1995;373:523–7.
21. Hsiao KK, Borchelt DR, Olson K, et al., Age-related CNS disorder and early death in transgenic FVB/N mice overexpressing Alzheimer amyloid precursor proteins, *Neuron*, 1995;15:1203–18.
22. Duff K, Eckman C, Zehr C, et al., Increased amyloid-beta42(43) in brains of mice expressing mutant presenilin 1, *Nature*, 1996;383:710–3.
23. McAllister C, Long J, Bowers A, et al., Genetic targeting aromatase in male amyloid precursor protein transgenic mice down-regulates beta-secretase (BACE1) and prevents Alzheimer-like pathology and cognitive impairment, *J Neurosci*, 2010;30:7326–34.
24. Borchelt DR, Thinakaran G, Eckman CB, et al., Familial Alzheimer's disease-linked presenilin 1 variants elevate Abeta1-42/1-40 ratio in vitro and in vivo, *Neuron*, 1996;17:1005–13.
25. Citron M, Westaway D, Xia W, et al., Mutant presenilins of Alzheimer's disease increase production of 42-residue amyloid beta-protein in both transfected cells and transgenic mice, *Nat Med*, 1997;3:67–72.
26. Borchelt DR, Ratovitski T, van Lare J, et al., Accelerated amyloid deposition in the brains of transgenic mice coexpressing mutant presenilin 1 and amyloid precursor proteins, *Neuron*, 1997;19:939–45.
27. Holcomb L, Gordon MN, McGowan E, et al., Accelerated Alzheimer-type phenotype in transgenic mice carrying both mutant amyloid precursor protein and presenilin 1 transgenes, *Nat Med*, 1998;4:97–100.
28. Oddo S, Caccamo A, Shepherd JD, et al., Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Abeta and synaptic dysfunction, *Neuron*, 2003;39:409–21.
29. Stalder M, Phinney A, Probst A, et al., Association of microglia with amyloid plaques in brains of APP23 transgenic mice, *Am J Pathol*, 1999;154:1673–84.
30. Wegiel J, Wang KC, Imaki H, et al., The role of microglial cells and astrocytes in fibrillar plaque evolution in transgenic APP(SW) mice, *Neurobiol Aging*, 2001;22:49–61.
31. Jin JJ, Kim HD, Maxwell JA, et al., Toll-like receptor 4-dependent upregulation of cytokines in a transgenic mouse model of Alzheimer's disease, *J Neuroinflammation*, 2008;5:23.
32. Grommes C, Lee CY, Wilkinson BL, et al., Regulation of microglial phagocytosis and inflammatory gene expression by Gas6 acting on the Axl/Mer family of tyrosine kinases, *J Neuroimmune Pharmacol*, 2008;3:130–40.
33. Sly LM, Krzesicki RF, Brashler JR, et al., Endogenous brain cytokine mRNA and inflammatory responses to lipopolysaccharide are elevated in the Tg2576 transgenic mouse model of Alzheimer's disease, *Brain Res Bull*, 2001;56:581–8.
34. Mori T, Koyama N, Arendash GW, et al., Overexpression of human S100B exacerbates cerebral amyloidosis and gliosis in the Tg2576 mouse model of Alzheimer's disease, *Glia*, 2010;58(3):300–14.
35. Tan J, Town T, Crawford F, et al., Role of CD40 ligand in amyloidosis in transgenic Alzheimer's mice, *Nat Neurosci*, 2002;5:1288–93.
36. Yamamoto M, Kiyota T, Horiba M, et al., Interferon-gamma and tumor necrosis factor-alpha regulate amyloid-beta plaque deposition and beta-secretase expression in Swedish mutant APP transgenic mice, *Am J Pathol*, 2007;170:680–92.
37. Fuhrmann M, Bittner T, Jung CK, et al., Microglial Cx3cr1 knockout prevents neuron loss in a mouse model of Alzheimer's disease, *Nat Neurosci*, 2010;13:411–3.
38. Lim GP, Yang F, Chu T, et al., Ibuprofen suppresses plaque pathology and inflammation in a mouse model for Alzheimer's disease, *J Neurosci*, 2000;20:5709–14.
39. Jantzen PT, Connor KE, DiCarlo G, et al., Microglial activation and beta-amyloid deposit reduction caused by a nitric oxide-releasing nonsteroidal anti-inflammatory drug in amyloid precursor protein plus presenilin-1 transgenic mice, *J Neurosci*, 2002;22:2246–54.
40. Yan Q, Zhang J, Liu H, et al., Anti-inflammatory drug therapy alters beta-amyloid processing and deposition in an animal model of Alzheimer's disease, *J Neurosci*, 2003;23:7504–9.
41. McKee AC, Carreras I, Hossain L, et al., Ibuprofen reduces Abeta, hyperphosphorylated tau and memory deficits in Alzheimer mice, *Brain Res*, 2008;1207:225–36.
42. Van Dam D, Coen K, De Deyn PP, Ibuprofen modifies cognitive disease progression in an Alzheimer's mouse model, *J Psychopharmacol*, 2010;24:383–8.
43. Choi JK, Jenkins BG, Carreras I, et al., Anti-inflammatory treatment in AD mice protects against neuronal pathology, *Exp Neurol*, 2010;223:377–84.
44. Wisniewski KE, Dalton AJ, McLachlan C, et al., Alzheimer's disease in Down's syndrome: clinicopathologic studies, *Neurology*, 1985;35:957–61.
45. Wisniewski KE, Wisniewski HM, Wen GY, Occurrence of neuropathological changes and dementia of Alzheimer's disease in Down's syndrome, *Ann Neurol*, 1985;17:278–82.
46. Tanzi RE, Molecular genetics of Alzheimer's disease and the amyloid beta peptide precursor gene, *Ann Med*, 1989;21:91–4.
47. Patterson D, Gardiner K, Kao FT, et al., Mapping of the gene encoding the beta-amyloid precursor protein and its relationship to the Down syndrome region of chromosome 21, *Proc Natl Acad Sci U S A*, 1988;85:8266–70.
48. Prasher VP, Farrer MJ, Kessling AM, et al., Molecular mapping of Alzheimer-type dementia in Down's syndrome, *Ann Neurol*, 1998;43:380–3.
49. Rovelet-Lecrux A, Hannequin D, et al., APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy, *Brain*, 2006;129:2977–83.
50. Sleegers K, Brouwers N, Gijsels I, et al., APP duplication is sufficient to cause early onset Alzheimer's dementia with cerebral amyloid angiopathy, *Brain*, 2006;129:2977–83.
51. Oyama F, Cairns NJ, Shimada H, et al., Down's syndrome: up-regulation of beta-amyloid protein precursor and tau mRNAs and their defective coordination, *J Neurochem*, 1994;62:1062–6.
52. Beyreuther K, Pollwein P, Multhaup G, et al., Regulation and expression of the Alzheimer's beta/A4 amyloid protein precursor in health, disease, and Down's syndrome, *Ann N Y Acad Sci*, 1993;695:91–102.
53. Cheon MS, Dierssen M, Kim SH, et al., Protein expression of BACE1, BACE2 and APP in Down syndrome brains, *Amino Acids*, 2008;35:339–43.
54. Choi JH, Berger JD, Mazzella MJ, et al., Age-dependent dysregulation of brain amyloid precursor protein in the Ts65Dn Down syndrome mouse model, *J Neurochem*, 2009;110:1818–27.
55. Ledoux S, Bergeron C, Nalbantoglu J, et al., Polymerase chain reaction quantification of lymphoid amyloid precursor protein mRNAs in Alzheimer's disease and Down's syndrome, *Neurosci Lett*, 1995;193:137–9.
56. Griffin WS, Stanley LC, Ling C, et al., Brain interleukin 1 and S-100 immunoreactivity are elevated in Down syndrome and Alzheimer disease, *Proc Natl Acad Sci U S A*, 1989;86:7611–5.
57. Griffin WS, Sheng JG, McKenzie JE, et al., Life-long overexpression of S100beta in Down's syndrome: implications for Alzheimer pathogenesis, *Neurobiol Aging*,

- 1998;19:401–5.
58. Rozemuller JM, Eikelenboom P, Pals ST, et al., Microglial cells around amyloid plaques in Alzheimer's disease express leucocyte adhesion molecules of the LFA-1 family, *Neurosci Lett*, 1989;101:288–92.
 59. Rozemuller JM, Eikelenboom P, Stam FC, Role of microglia in plaque formation in senile dementia of the Alzheimer type. An immunohistochemical study, *Virchows Arch B Cell Pathol Incl Mol Pathol*, 1986;51:247–54.
 60. Griffin WS, Sheng JG, Roberts GW, et al., Interleukin-1 expression in different plaque types in Alzheimer's disease: significance in plaque evolution, *J Neuropathol Exp Neurol*, 1995;54:276–81.
 61. Sheng JG, Mrak RE, Griffin WS, Neuritic plaque evolution in Alzheimer's disease is accompanied by transition of activated microglia from primed to enlarged to phagocytic forms, *Acta Neuropathol (Berl)*, 1997;94:1–5.
 62. Sheng JG, Mrak RE, Griffin WS, Microglial interleukin-1 alpha expression in brain regions in Alzheimer's disease: correlation with neuritic plaque distribution, *Neuropathol Appl Neurobiol*, 1995;21:290–301.
 63. Sheng JG, Mrak RE, Griffin WS, Glial-neuronal interactions in Alzheimer disease: progressive association of IL-1alpha+ microglia and S100beta+ astrocytes with neurofibrillary tangle stages, *J Neuropathol Exp Neurol*, 1997;56:285–90.
 64. Sheng JG, Zhu SG, Jones RA, et al., Interleukin-1 promotes expression and phosphorylation of neurofilament and tau proteins in vivo, *Exp Neurol*, 2000;163:388–91.
 65. Sheng JG, Jones RA, Zhou XQ, et al., Interleukin-1 promotion of MAPK-p38 overexpression in experimental animals and in Alzheimer's disease: potential significance for tau protein phosphorylation, *Neurochem Int*, 2001;39:341–8.
 66. Mrak RE, Sheng JG, Griffin WS, Correlation of astrocytic S100 beta expression with dystrophic neurites in amyloid plaques of Alzheimer's disease, *J Neuropathol Exp Neurol*, 1996;55:273–9.
 67. Li Y, Liu L, Kang J, et al., Neuronal-glia interactions mediated by interleukin-1 enhance neuronal acetylcholinesterase activity and mRNA expression, *J Neurosci*, 2000;20:149–55.
 68. Nichol KE, Poon WW, Parachikova AI, et al., Exercise alters the immune profile in Tg2576 Alzheimer mice toward a response coincident with improved cognitive performance and decreased amyloid, *J Neuroinflammation*, 2008;5:13.
 69. Wu CK, Thal L, Pizzo D, et al., Apoptotic signals within the basal forebrain cholinergic neurons in Alzheimer's disease, *Exp Neurol*, 2005;195:484–96.
 70. Sheng JG, Mrak RE, Jones RA, et al., Neuronal DNA damage correlates with overexpression of interleukin-1beta converting enzyme in APPV717F mice, *Neurobiol Aging*, 2001;22:895–902.
 71. Zhu SG, Sheng JG, Jones RA, et al., Increased interleukin-1beta converting enzyme expression and activity in Alzheimer disease, *J Neuropathol Exp Neurol*, 1999;58:582–7.
 72. Chan SL, Griffin WS, Mattson MP, Evidence for caspase-mediated cleavage of AMPA receptor subunits in neuronal apoptosis and Alzheimer's disease, *J Neurosci Res*, 1999;57:315–23.
 73. Moncaster JA, Pineda R, Moir RD, et al., Alzheimer's disease amyloid-beta links lens and brain pathology in Down syndrome, *PLoS One*, 2010;5:e10659.
 74. Westmark CJ, Westmark PR, Malter JS, Alzheimer's disease and Down syndrome rodent models exhibit audiogenic seizures, *J Alzheimers Dis*, 2010;20:1009–13.
 75. Cataldo AM, Petanceska S, Peterhoff CM, et al., App gene dosage modulates endosomal abnormalities of Alzheimer's disease in a segmental trisomy 16 mouse model of down syndrome, *J Neurosci*, 2003;23:6788–92.
 76. Chen Y, Dyakin VV, Branch CA, et al., In vivo MRI identifies cholinergic circuitry deficits in a Down syndrome model, *Neurobiol Aging*, 2009;30:1453–65.
 77. Salehi A, Delcroix JD, Belichenko PV, et al., Increased App expression in a mouse model of Down's syndrome disrupts NGF transport and causes cholinergic neuron degeneration, *Neuron*, 2006;51:29–42.
 78. Jiang Y, Mullaney KA, Peterhoff CM, et al., Alzheimer's-related endosome dysfunction in Down syndrome is Abeta-independent but requires APP and is reversed by BACE-1 inhibition, *Proc Natl Acad Sci U S A*, 2010;107:1630–5.
 79. Fukutani Y, Cairns NJ, Rossor MN, et al., Cerebellar pathology in sporadic and familial Alzheimer's disease including APP 717 (Val→Ile) mutation cases: a morphometric investigation, *J Neural Sci*, 1997;149:177–84.
 80. Sheng JG, Mrak RE, Griffin WS, Progressive neuronal DNA damage associated with neurofibrillary tangle formation in Alzheimer disease, *J Neuropathol Exp Neurol*, 1998;57:323–8.
 81. Wirths O, Beyrehan H, Marcelllo A, et al., Inflammatory changes are tightly associated with neurodegeneration in the brain and spinal cord of the APP/PS1 mouse model of Alzheimer's disease, *Neurobiol Aging*, 2010;31:747–57.
 82. Mrak RE, Neuropathology and the neuroinflammation idea, *J Alzheimers Dis*, 2009;18:473–81.
 83. Burger PC, Vogel FS, The development of the pathologic changes of Alzheimer's disease and senile dementia in patients with Down's syndrome, *Am J Pathol*, 1973;73:457–76.
 84. Royston MC, McKenzie JE, Gentleman SM, et al., Overexpression of S100beta in Down's syndrome: correlation with patient age and with beta-amyloid deposition, *Neuropathol Appl Neurobiol*, 1999;25:387–93.
 85. Mann DM, Royston MC, Ravindra CR, Some morphometric observations on the brains of patients with Down's syndrome: their relationship to age and dementia, *J Neurol Sci*, 1990;99:153–64.
 86. Goldgaber D, Harris HW, Hla T, et al., Interleukin 1 regulates synthesis of amyloid beta-protein precursor mRNA in human endothelial cells, *Proc Natl Acad Sci U S A*, 1989;86:7606–10.
 87. Li Y, Wang J, Sheng JG, et al., S100 beta increases levels of beta-amyloid precursor protein and its encoding mRNA in rat neuronal cultures, *J Neurochem*, 1998;71:1421–8.
 88. Mrak RE, Griffin WS, The role of activated astrocytes and of the neurotrophic cytokine S100B in the pathogenesis of Alzheimer's disease, *Neurobiol Aging*, 2001;22:915–22.
 89. Sheng JG, Griffin WS, Royston MC, et al., Distribution of interleukin-1-immunoreactive microglia in cerebral cortical layers: implications for neuritic plaque formation in Alzheimer's disease, *Neuropathol Appl Neurobiol*, 1998;24:278–83.
 90. Tanzi RE, McClatchey AJ, Lamperti ED, et al., Protease inhibitor domain encoded by an amyloid protein precursor mRNA associated with Alzheimer's disease, *Nature*, 1988;331:528–30.
 91. Barger SW, Harmon AD, Microglial activation by Alzheimer amyloid precursor protein and modulation by apolipoprotein E, *Nature*, 1997;388:878–81.
 92. Griffin WS, Excess betaCTF, not Abeta: the culprit in Alzheimer-related endocytic dysfunction, *Proc Natl Acad Sci U S A*, 2010;107:1263–4.
 93. Ghosal K, Stathopoulos A, Pimplikar SW, APP intracellular domain impairs adult neurogenesis in transgenic mice by inducing neuroinflammation, *PLoS One*, 2010;5:e11866.
 94. Goshen I, Kreisel T, Ben-Menachem-Zidon O, et al., Brain interleukin-1 mediates chronic stress-induced depression in mice via adrenocortical activation and hippocampal neurogenesis suppression, *Mol Psychiatry*, 2008;13:717–28.
 95. Koo JW, Duman RS, IL-1beta is an essential mediator of the antineurogenic and anhedonic effects of stress, *Proc Natl Acad Sci U S A*, 2008;105:751–6.
 96. Sheng JG, Mrak RE, Bales KR, et al., Overexpression of the neurotrophic cytokine S100beta precedes the appearance of neuritic beta-amyloid plaques in APPV717F mice, *J Neurochem*, 2000;74:295–301.
 97. Heneka MT, Sastre M, Dumitrescu-Ozimek L, et al., Focal glial activation coincides with increased BACE1 activation and precedes amyloid plaque deposition in APPV717F transgenic mice, *J Neuroinflammation*, 2005;2:22.
 98. Chen Q, Nakajima A, Choi SH, et al., Loss of presenilin function causes Alzheimer's disease-like neurodegeneration in the mouse, *J Neurosci Res*, 2008;86:1615–25.
 99. El Khoury J, Toft M, Hickman SE, et al., Ccr2 deficiency impairs microglial accumulation and accelerates progression of Alzheimer-like disease, *Nat Med*, 2007;13:432–8.
 100. Grathwohl SA, Kalin RE, Bolmont T, et al., Formation and maintenance of Alzheimer's disease beta-amyloid plaques in the absence of microglia, *Nat Neurosci*, 2009;12:1361–3.
 101. Akiyama H, Barger S, Barnum S, et al., Inflammation and Alzheimer's disease, *Neurobiol Aging*, 2000;21:383–421.
 102. Fonseca MI, Ager RR, Chu SH, et al., Treatment with a C5aR antagonist decreases pathology and enhances behavioral performance in murine models of Alzheimer's disease, *J Immunol*, 2009;183:1375–83.
 103. Bonifati DM, Kishore U, Role of complement in neurodegeneration and neuroinflammation, *Mol Immunol*, 2007;44(5):999–1010.
 104. Corder EH, Saunders AM, Strittmatter WJ, et al., Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families, *Science*, 1993;261:921–3.
 105. Roses AD, Saunders AM, Corder EH, et al., Influence of the susceptibility genes apolipoprotein E-epsilon 4 and apolipoprotein E-epsilon 2 on the rate of disease expressivity of late-onset Alzheimer's disease, *Arzmittelforschung*, 1995;45:413–7.
 106. Sheng JG, Mrak RE, Griffin WS, Apolipoprotein E distribution among different plaque types in Alzheimer's disease: implications for its role in plaque progression, *Neuropathol Appl Neurobiol*, 1996;22:334–41.
 107. Drzezgza A, Grimmer T, Henriksen G, et al., Effect of APOE genotype on amyloid plaque load and gray matter volume in Alzheimer disease, *Neurology*, 2009;72:1487–94.
 108. Vemuri P, Wiste HJ, Weigand SD, et al., Effect of apolipoprotein E on biomarkers of amyloid load and neuronal pathology in Alzheimer disease, *Ann Neurol*, 2010;67:308–16.
 109. Chen Y, Durakoglugil MS, Xian X, et al., ApoE4 reduces glutamate receptor function and synaptic plasticity by selectively impairing ApoE receptor recycling, *Proc Natl Acad Sci U S A*, 2010;107:12011–6.
 110. Barger SW, DeWall KM, Liu L, et al., Relationships between expression of apolipoprotein E and beta-amyloid precursor protein are altered in proximity to Alzheimer beta-amyloid plaques; potential explanations from cell culture studies, *J Neuropathol Exp Neurol*, 2008;67(8):773–83.
 111. Egenstepberger R, Kosel S, von Eitzen U, et al., Microglial activation in Alzheimer disease: Association with APOE genotype, *Brain Pathol*, 1998;8:439–47.
 112. In t'Veld BA, Ruitenbergh A, Hofman A, et al., Nonsteroidal antiinflammatory drugs and the risk of Alzheimer's disease, *N Engl J Med*, 2001;345:1515–21.
 113. Cornelius C, Fastbom J, Winblad B, et al., Aspirin, NSAIDs, risk of dementia, and influence of the apolipoprotein E epsilon 4 allele in an elderly population, *Neuroepidemiology*, 2004;23:135–43.
 114. Yip AG, Green RC, Huyck M, et al., Nonsteroidal anti-inflammatory drug use and Alzheimer's disease risk: the MIRAGE Study, *BMC Geriatr*, 2005;5:2.
 115. Hayden KM, Zandi PP, Khachaturian AS, et al., Does NSAID use modify cognitive trajectories in the elderly? The Cache County study, *Neurology*, 2007;69:275–82.
 116. Szekeley CA, Bretiner JC, Fitzpatrick AL, et al., NSAID use and dementia risk in the Cardiovascular Health Study: role of APOE and NSAID type, *Neurology*, 2008;70:17–24.
 117. Scharf S, Mander A, Ugoni A, et al., A double-blind, placebo-controlled trial of diclofenac/misoprostol in Alzheimer's disease, *Neurology*, 1999;53:197–201.
 118. Aisen PS, Schafer KA, Grundman M, et al., Effects of rofecoxib or naproxen vs placebo on Alzheimer disease progression: a randomized controlled trial, *JAMA*, 2003;289:2819–26.
 119. Reines SA, Block GA, Morris JC, et al., Rofecoxib: no effect on Alzheimer's disease in a 1-year, randomized, blinded, controlled study, *Neurology*, 2004;62:66–71.
 120. Pasqualetti P, Bonomini C, Dal Forno G, et al., A randomized controlled study on effects of ibuprofen on cognitive progression of Alzheimer's disease, *Acta Neurol Scand*, 2009;121:102–10.
 121. Colton CA, Brown CM, Cook D, et al., APOE and the regulation of microglial nitric oxide production: a link between genetic risk and oxidative stress, *Neurobiol Aging*, 2002;23:777–85.
 122. Ophir G, Amariglio N, Jacob-Hirsch J, et al., Apolipoprotein E4 enhances brain inflammation by modulation of the NF-kappaB signaling cascade, *Neurobiol Dis*, 2005;20:709–18.
 123. Chen S, Averett NT, Manelli A, et al., Isoform-specific effects of apolipoprotein E on secretion of inflammatory mediators in adult rat microglia, *J Alzheimers Dis*, 2005;7:25–35.
 124. Biffi A, Anderson CD, Desikan RS, et al., Genetic variation and neuroimaging measures in Alzheimer disease, *Arch Neurol*, 2010;67:677–85.
 125. Andrawis JP, Hwang KS, Green AE, et al., Effects of ApoE4 and maternal history of dementia on hippocampal atrophy, *Neurobiol Aging*, 2010; [Epub ahead of print].
 126. Ho AJ, Raji CA, Becker JT, et al., Obesity is linked with lower brain volume in 700 AD and MCI patients, *Neurobiol Aging*, 2010;31:1326–39.
 127. Apostolova LG, Hwang KS, Andrawis JP, et al., 3D PIB and CSF biomarker associations with hippocampal atrophy in ADNI subjects, *Neurobiol Aging*, 2010;31:1284–303.
 128. Nettiksimmons J, Harvey D, Brewer J, et al., Subtypes based on cerebrospinal fluid and magnetic resonance imaging markers in normal elderly predict cognitive decline, *Neurobiol Aging*, 2010;31:1419–28.
 129. Iwatsubo T, Japanese Alzheimer's Disease Neuroimaging Initiative: present status and future, *Alzheimers Dement*, 2010;6:297–9.
 130. Di Bona D, Candore G, Franceschi C, et al., Systematic review by meta-analyses on the possible role of TNF-alpha polymorphisms in association with Alzheimer's disease, *Brain Res Rev*, 2009;61:160–8.
 131. Vural P, Degirmencioglu S, Parildar-Karpuzoglu H, et al., The combinations of TNFalpha-308 and IL-6 -174 or IL-10 -1082 genes polymorphisms suggest an association with susceptibility to sporadic late-onset Alzheimer's disease, *Acta Neurol Scand*, 2009;120:396–401.
 132. Nicoll JA, Mrak RE, Graham DI, et al., Association of interleukin-1 gene polymorphisms with Alzheimer's disease, *Ann Neurol*, 2000;47:365–8.
 133. Rebeck GW, Confirmation of the genetic association of interleukin-1A with early onset sporadic Alzheimer's disease, *Neurosci Lett*, 2000;293:75–7.
 134. Grimaldi LM, Casadei VM, Ferri C, et al., Association of early-onset Alzheimer's disease with an interleukin-1alpha gene polymorphism, *Ann Neurol*, 2000;47:361–5.
 135. Licastro F, Veglia F, Chiappelli M, et al., A polymorphism of the interleukin-1 beta gene at position +3953 influences progression and neuro-pathological hallmarks of Alzheimer's disease, *Neurobiol Aging*, 2004;25:1017–22.
 136. Sciacca FL, Ferri C, Licastro F, et al., Interleukin-1B polymorphism is associated with age at onset of Alzheimer's disease, *Neurobiol Aging*, 2003;24:927–31.
 137. Seripa D, Matera MG, Dal Forno G, et al., Genotypes and haplotypes in the IL-1 gene cluster: analysis of two genetically and diagnostically distinct groups of Alzheimer patients, *Neurobiol Aging*, 2005;26:455–64.
 138. Hu JL, Li G, Zhou DX, et al., Genetic analysis of interleukin-1A C(-889)T polymorphism with Alzheimer disease, *Cell Mol Neurobiol*, 2009;29:81–5.
 139. Tsai SJ, Liu HC, Liu TY, et al., Lack of association between the interleukin-1alpha gene C(-889)T polymorphism and Alzheimer's disease in a Chinese population, *Neurosci Lett*, 2003;343:93–6.
 140. Déniz-Naranjo MC, Muñoz-Fernandez C, Alemany-Rodríguez MJ, et al., Cytokine IL-1 beta but not IL-1 alpha promoter polymorphism is associated with Alzheimer disease in a population from the Canary Islands, Spain, *Eur J Neurol*, 2008;15:1080–4.
 141. Di Bona D, Plaia A, Vasto S, et al., Association between the interleukin-1beta polymorphisms and Alzheimer's disease: a systematic review and meta-analysis, *Brain Res Rev*, 2008;59:155–63.
 142. Klimkowicz-Mrowiec A, Marona M, Wolkow P, et al., Interleukin-1 gene -511 CT polymorphism and the risk of

- Alzheimer's disease in a Polish population, *Dement Geriatr Cogn Disord*, 2009;28:461-4.
143. Ravaglia G, Paola F, Maioli F, et al., Interleukin-1beta and interleukin-6 gene polymorphisms as risk factors for AD: a prospective study, *Exp Gerontol*, 2006;41:85-92.
 144. Rainero I, Bo M, Ferrero M, et al., Association between the interleukin-1alpha gene and Alzheimer's disease: a meta-analysis, *Neurobiol Aging*, 2004;25:1293-8.
 145. Combarros O, Llorca J, Sanchez-Guerra M, et al., Age-dependent association between interleukin-1A (-889) genetic polymorphism and sporadic Alzheimer's disease. A meta-analysis, *J Neurol*, 2003;250:987-9.
 146. Combarros O, Sanchez-Guerra M, Infante J, et al., Gene dose-dependent association of interleukin-1A -889 allele 2 polymorphism with Alzheimer's disease, *J Neurol*, 2002;249:1242-5.
 147. Yucesoy B, Peila R, White LR, et al., Association of interleukin-1 gene polymorphisms with dementia in a community-based sample: the Honolulu-Asia Aging Study, *Neurobiol Aging*, 2006;27:211-7.
 148. Bertram L, McQueen MB, Mullin K, et al., Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database, *Nat Genet*, 2007;39:17-23.
 149. Ribizzi G, Fiordoro S, Barocci S, et al., Cytokine polymorphisms and Alzheimer disease: possible associations, *Neuro Sci*, 2010;31:321-5.
 150. Du Y, Dodel RC, Eastwood BJ, et al., Association of an interleukin 1 alpha polymorphism with Alzheimer's disease, *Neurology*, 2000;55:480-3.
 151. Hedley R, Hallmayer J, Groth DM, et al., Association of interleukin-1 polymorphisms with Alzheimer's disease in Australia, *Ann Neurol*, 2002;51:795-7.
 152. Mrak RE, Griffin WS, Gila and their cytokines in progression of neurodegeneration, *Neurobiol Aging*, 2005;26:349-54.
 153. Mackenzie IR, Senile plaques do not progressively accumulate with normal aging, *Acta Neuropathol (Berl)*, 1994;87:520-5.
 154. Sperling RA, Laviolette PS, O'Keefe K, et al., Amyloid deposition is associated with impaired default network function in older persons without dementia, *Neuron*, 2009;63:178-88.
 155. Mackenzie IR, Hao C, Munoz DG, Role of microglia in senile plaque formation, *Neurobiol Aging*, 1995;16:797-804.
 156. Sheng JG, Mrak RE, Rovnaghi CR, et al., Human brain S100 beta and S100 beta mRNA expression increases with age: pathogenic implications for Alzheimer's disease, *Neurobiol Aging*, 1996;17:359-63.
 157. Sheng JG, Mrak RE, Griffin WS, Enlarged and phagocytic, but not primed, interleukin-1 alpha-immunoreactive microglia increase with age in normal human brain, *Acta Neuropathol*, 1998;95:229-34.
 158. Gouras GK, Relkin NR, Sweeney D, et al., Increased apolipoprotein E epsilon 4 in epilepsy with senile plaques, *Ann Neurol*, 1997;41:402-4.
 159. Teasdale GM, Murray GD, Nicoll JA, The association between APOE epsilon4, age and outcome after head injury: a prospective cohort study, *Brain*, 2005;128:2556-61.
 160. Sparks DL, Hunsaker JC, 3rd, Scheff SW, et al., Cortical senile plaques in coronary artery disease, aging and Alzheimer's disease, *Neurobiol Aging*, 1990;11:601-7.
 161. Esiri MM, Biddolph SC, Morris CS, Prevalence of Alzheimer plaques in AIDS, *J Neurol Neurosurg Psychiatry*, 1998;65:29-33.
 162. Stanley LC, Mrak RE, Woody RC, et al., Glial cytokines as neuropathogenic factors in HIV infection: pathogenic similarities to Alzheimer's disease, *J Neuropathol Exp Neurol*, 1994;53:231-8.
 163. Biessels GJ, Staekenborg S, Brunner E, et al., Risk of dementia in diabetes mellitus: a systematic review, *Lancet Neurol*, 2006;5:64-74.
 164. Dore GA, Elias MF, Robbins MA, et al., Presence of the APOE epsilon4 allele modifies the relationship between type 2 diabetes and cognitive performance: the Maine-Syracuse Study, *Diabetologia*, 2009;52(12):2551-60.
 165. Xu WL, von Strauss E, Qiu CX, et al., Uncontrolled diabetes increases the risk of Alzheimer's disease: a population-based cohort study, *Diabetologia*, 2009;52:1031-9.
 166. Kloppenborg RP, van den Berg E, Kappelle LJ, et al., Diabetes and other vascular risk factors for dementia: which factor matters most? A systematic review, *Eur J Pharmacol*, 2008;585:97-108.
 167. van den Berg E, Kloppenborg RP, Kessels RP, et al., Type 2 diabetes mellitus, hypertension, dyslipidemia and obesity: A systematic comparison of their impact on cognition, *Biochim Biophys Acta*, 2009;1792:470-81.
 168. Xu WL, Qiu CX, Wahlin A, et al., Diabetes mellitus and risk of dementia in the Kungsholmen project: a 6-year follow-up study, *Neurology*, 2004;63:1181-6.
 169. Yoshitake T, Kiyohara Y, Kato I, et al., Incidence and risk factors of vascular dementia and Alzheimer's disease in a defined elderly Japanese population: the Hisayama Study, *Neurology*, 1995;45:1161-8.
 170. Peila R, White LR, Petrovich H, et al., Joint effect of the APOE gene and midlife systolic blood pressure on late-life cognitive impairment: the Honolulu-Asia aging study, *Stroke*, 2001;32:2882-9.
 171. Peila R, Rodriguez BL, Launer LJ, Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: The Honolulu-Asia Aging Study, *Diabetes*, 2002;51:1256-62.
 172. Griffin WS, Sheng JG, Gentleman SM, et al., Microglial interleukin-1 alpha expression in human head injury: correlations with neuronal and neuritic beta-amyloid precursor protein expression, *Neurosci Lett*, 1994;176:133-6.
 173. Gentleman SM, Leclercq PD, Moyes L, et al., Long-term intracerebral inflammatory response after traumatic brain injury, *Forensic Sci Int*, 2004;146:97-104.
 174. Graham DI, Gentleman SM, Nicoll JA, et al., Altered beta-APP metabolism after head injury and its relationship to the aetiology of Alzheimer's disease, *Acta Neurochir Suppl*, 1996;66:96-102.
 175. Griffin WS, Mrak RE, Interleukin-1 in the genesis and progression of and risk for development of neuronal degeneration in Alzheimer's disease, *J Leukoc Biol*, 2002;72:233-8.
 176. Griffin WS, Yeralan O, Sheng JG, et al., Overexpression of the neurotrophic cytokine S100 beta in human temporal lobe epilepsy, *J Neurochem*, 1995;65:228-33.
 177. Sheng JG, Boop FA, Mrak RE, Griffin WS, Increased neuronal beta-amyloid precursor protein expression in human temporal lobe epilepsy: association with interleukin-1 alpha immunoreactivity, *J Neurochem*, 1994;63:1872-79.
 178. Wisniewski TL, Mendel E, Morse EV, et al., Relationship between serum cyclo(His-Pro) concentrations and the nutritional status of HIV-infected patients, *South Med J*, 1994;87:348-51.